

PASADENA CYTO-PATHOLOGY LABORATORY, INC.

A MEDICAL CORPORATION

100 W. CALIFORNIA BLVD

PASADENA, CA 91105

Billing Inquiries: (626)397-3448

S. S. Murakami, M.D., Director (CLIA)

Tissue Information: (626)397-8616

H. D. Slosser, M. D., Director

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CYTOLOGY SPECIMEN COLLECTION PROCEDURES

All slides and specimen containers must be labeled with patient name and specimen source and accompanied by a completed requisition

TEST NAME	SPECIMEN REQUIREMENTS	SPECIAL INSTRUCTIONS/STORAGE	Result availability from time received in the Lab
FINE NEEDLE ASPIRATION	Needle aspirate specimen in Cytolyt Solution; prepared slides.	Expel aspirated material directly into Cytolyt Solution. Use a new needle and syringe for each pass. Smears: prepare 2 slides per pass. Expel any additional material into the Cytolyt solution. Label frosted-end slides with the patient's name. Be sure to put the aspirated material on the right side of the slide (you should be able to read the word "Specimen"). Smear two slides together, then immediately spray with Cytology fixative. Allow fixative to dry before placing in plastic containers for transport. Refrigeration of specimens in Cytolyt is not necessary. Excessive blood may limit interpretation. A negative or non-diagnostic report may reflect poor needle placement and does not completely rule out a malignancy. If multiple lab tests are ordered, collect a separate, unfixed specimen for other tests.	24 to 48 hours
URINARY TRACT SPECIMENS	Hydrate patient before obtaining specimen. SPECIMEN REQUIREMENTS: Voided or catheterized urine. MINIMAL VOLUME: At least 50ML is optimal for voided specimens. Submit specimen fresh in a non-sterile, leakproof container, or in Cytolyt Solution.	Label container with source of specimen. Contact PCL for specimen pick-up as soon as possible after collection. Refrigerate the specimen if there is any delay in specimen pick-up, or if the specimen must sit overnight. Specimens collected in Cytolyt Solution do not need refrigeration. Specimens collected in Urocyte for FISH studies must be refrigerated and sent to PCL as soon as possible after collection (see procedure for FISH). Improper fixation, 24-hour collection or undue delay in transport will render a specimen unsatisfactory for evaluation. Calculi, recent instrumentation, and chemotherapy may produce atypical changes in urothelial cells simulating malignancy. When microbiology tests are also ordered, collect a separate specimen for microbiology in a sterile container.	24 to 48 hours
CSF	Fresh, unfixed spinal fluid. Not less than 1 ml.	Do not freeze. Contact PCL for specimen pick-up as soon as possible after collection. Refrigerate the specimen if there is any delay in specimen pick-up, or if the specimen must sit overnight. Diagnostic and/or therapeutic procedures should be noted on the requisition as they can produce striking cytologic changes.	24 to 48 hours

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BODY FLUID SPECIMENS Pleural fluid, thoracentesis fluid, peritoneal fluid, ascitic fluid, paracentesis fluid, pericardial fluid, pericardiocentesis fluid, synovial fluid, culdocentesis fluid or peritoneal washing.	MINIMAL VOLUME: 5 ML fresh, unfixed body fluid. Body fluid specimens for <u>cytology only</u> may be collected in Cytolyt Solution.	For shared specimens with Microbiology, specimen must be sterile. Label container with source of specimen. Contact PCL for specimen pick-up as soon as possible after collection. Refrigerate the specimen if there is any delay in specimen pick-up, or if the specimen must sit overnight.	24 to 48 hours
BRONCHIAL WASH/BRUSH	Prepared slides and/or brush from lesion area and/or saline washing of bronchus. Brushing-2 prepared slides in Pap fixative or brush tip in Preservcyt Solution. Washing-not less than 2 ML	For shared specimens with Microbiology, specimen must be sterile. Refrigerate the specimen if there is any delay in specimen pick-up, or if the specimen must sit overnight. For site-selective cytology studies, each specimen must be separate, source identified, and submitted with a separate requisition.	24 to 48 hours
BREAST NIPPLE SECRETION	Secretion from breast nipple may be submitted for cytologic evaluation.	Hold slide to nipple and smear secretion across slide. Fix immediately with spray fixative, allow to dry and place slide in a plastic or cardboard slide holder for transport.	24 to 48 hours
SPUTUM	Spontaneous or induced sputum may be submitted to cytology for evaluation. The material submitted for cytologic evaluation of the lung should be a deep-cough specimen. Saliva or nasal secretions are not adequate for this purpose. At least 3ml (one tablespoon) is optimal. Specimen may be collected in Cytolyt for <u>cytology only</u> .	For shared specimens with Microbiology, specimen must be sterile. Label container with source of specimen. Contact PCL for specimen pick-up as soon as possible after collection. Refrigerate the specimen if there is any delay in specimen pick-up, or if the specimen must sit overnight. Gross contamination of specimen by food particles limits evaluation. 24-hour sputum collection for cytology is unacceptable due to excessive bacterial overgrowth. Absence of pulmonary macrophages will result in specimen being reported as unsatisfactory for evaluation.	24 to 48 hours

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TEST NAME	SPECIMEN REQUIREMENTS	SPECIAL INSTRUCTIONS/STORAGE	Result availability from time received in the Lab
ENDOSCOPY SPECIMENS	Specimens collected during endoscopy may include the following: esophageal brushings/washings/biopsies, gastric brushings/washings/biopsies, washings/brushings/biopsies of small or large intestines and associated ducts/glands.	Label container with source of specimen. Contact PCL for specimen pick-up as soon as possible after collection. Refrigerate the specimen if there is any delay in specimen pick-up, or if the specimen must sit overnight. Gastric washing specimens are usually sub-optimal or unsatisfactory due to rapid degeneration of cellular material. Allowing smears to dry before they are fixed will render them unsatisfactory for cytologic evaluation. PROCEDURE FOR PREPARED SLIDES: When prepared at the time of endoscopy, immediate fixation is essential. Have the alcohol fixative bottles open and ready before any smears are prepared. Label the end of the slide with the patient's complete name. Roll the brush gently across the surface of the slide and place immediately into the fixative bottle. Prepare 2 slides from each site and place back-to-back in the fixative bottle. PROCEDURE FOR BRUSH: Do not put the brush in the alcohol bottle with the slides. Snip off brush and submit in a specimen container of saline or Cytolyt Solution. PROCEDURE FOR WASHINGS: Do not prepare slides. Submit fresh specimen in a sealable, leak-proof container and transport immediately to the laboratory. PROCEDURE FOR TISSUE BIOPSY: Put in a container of formalin fixative. Label container and complete a general requisition.	24 to 48 hours

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TEST NAME	SPECIMEN REQUIREMENTS	SPECIAL INSTRUCTIONS/STORAGE	Result availability from time received in the Lab
GYNECOLOGIC SPECIMENS (PAP SMEARS)	Specimens collected from various sites in the female genital tract may be submitted for cytologic evaluation (pap smears). Specimens may be collected from the vagina, cervix, endocervix and/or endometrium.	<p>Thin Prep: Do not use lubricant. Rinse collection device (spatula, brush or "broom") as quickly as possible. For brush: use a swirling motion while pressing the brush against the side of the collection vial. For broom: press the broom against the bottom of the vial 10 times, forcing the bristles apart, then swirl the broom vigorously in the collection vial. Discard the collection device. Refrigeration is unnecessary.</p> <p>High Risk HPV/Molecular Studies: High risk HPV and/or Gonorrhea/Chlamydia detection studies may be requested on specimens collected in Thin Prep vials. These studies are an important adjunct to the management of women with a diagnosis of ASC-US. Physicians may contract with PCL for reflexive HPV studies and designate the desired threshold for testing. Physicians may also request HPV and/or Molecular studies on a case-by -case basis by checking the appropriate box on the specimen requisition. The stability range for HPV/Molecular studies is 28 days from collection.</p> <p>SPECIMEN LABELING: Label the Thin Prep Vial with the patient's full name, specimen source and date of collection. A completed requisition must accompany each specimen.</p> <p>CLINICAL DATA: include any pertinent clinical data and/or patient history on the requisition. Include date of last menstrual period if applicable. LIMITATIONS: Inadequate cell sample limits evaluation. Lubricant use may render a sample unsatisfactory for evaluation. Excessive inflammation may limit evaluation. Insufficient patient history may limit evaluation.</p>	48 to 72 hours after receipt